Total Synthesis of Patulolide C and Its Homo, Nor, and Iso Analogs

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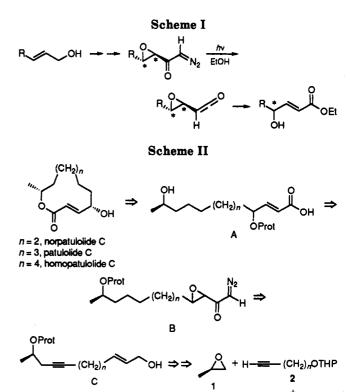
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The stereospecific total synthesis of the naturally occurring macrolide patulolide C 19Eb as well as its iso, nor, and homo analogs is described by applying the photoinduced rearrangement of enantiomerically pure epoxy diazomethyl ketones 14 to γ -hydroxy α,β -unsaturated esters 15 as the key step. The required epoxy diazomethyl ketones 14 are obtained by a Sharpless epoxidation of an appropriate allylic alcohol, followed by ruthenium tetraoxide oxidation to an oxiranecarboxylic acid, conversion into a mixed anhydride, and treatment with diazomethane. Macrolide 19Zb, which is a geometrical isomer of 19Eb, turned out to be a diastereomer of natural macrolide isopatulolide C, which implies the 4R,11R configuration for this natural material. X-ray diffraction analyses of 19Ea and 19Eb show that there is a considerable difference in spatial arrangement; particularly, the different torsion angles between the carbonyl and olefinic bonds are noteworthy. The conformational behavior of these macrolides is also deduced from the NMR and UV spectra.

In our study of the chemistry of functionalized epoxides, we showed that epoxy diazomethyl ketones undergo an interesting photoinduced rearrangement leading to γ -hydroxy α,β -unsaturated esters.^{1,2} Initially, an epoxy ketene is being formed,¹ which subsequently reacts with an alcoholic solvent to give a hydroxy alkene ester (Scheme I). With optically active substrates, which are readily accessible from allylic alcohols using the Sharpless epoxidation and subsequent oxidation to the corresponding oxiranecarboxylic acids, the γ -hydroxy function in these alkene esters can be introduced in an enantiocontrolled fashion.² The conversion of allylic alcohols into γ -hydroxy alkene esters is of synthetic value as was demonstrated by the total synthesis of various naturally occurring macrocyclic lactones such as aspicilin,³ colletallol,⁴ and pyrenophorol⁵ and the macrocyclic subunit of cytochalasan B.⁶

In this paper, we describe the total synthesis⁷ of patulolide C, a macrolide that has been isolated from the culture broth of *Penicillium urticae*.^{8,9} In addition, the synthesis of the 11-membered ring (norpatulolide C) and 13-membered ring (homopatulolide C) analogs as well as the corresponding Z isomers of these three patulolides will be reported. The retrosynthesis of patulolide C and its nor and homo analogs is outlined in Scheme II. Delactonization leads to diol alkene acid A which in principle is accessible via epoxy diazomethyl ketone B by applying the chemistry shown in Scheme I. The required allylic alcohol C can be built up from methyloxirane, an acetylenic alcohol, and a suitable two-carbon synthon. The involvement of enantiomerically pure methyloxirane en-

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sures the introduction of the chiral center adjacent to the lactone oxygen.

2-carbon synthon

Results and Discussion

The actual synthesis of the epoxy diazomethyl ketones **B** is shown in Scheme III. The alkynols, viz 4-pentyn-1-ol,¹⁰ 5-hexyn-1-ol,¹¹ and 6-heptyn-1-ol,¹² required for the opening of (R)-(+)-methyloxirane¹³ (1) were prepared by known procedures. The secondary alcohol function obtained in this ring-opening reaction was protected as a benzyl ether.¹⁴ Chain elongation of compounds **5** was achieved by oxidizing the primary alcohol of **5** to an

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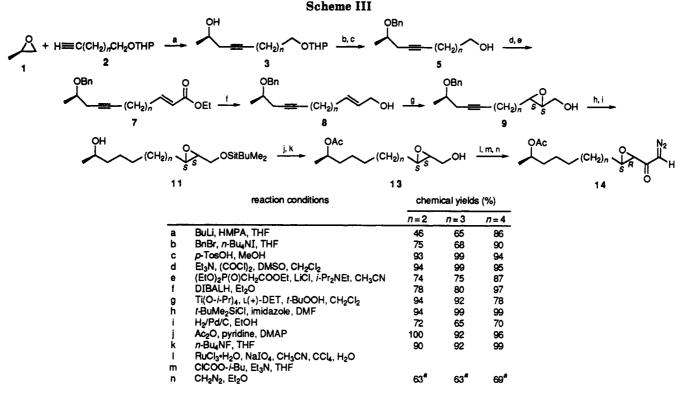
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^a Yields after step l, m, and n.

aldehyde group using the Swern oxidation and a subsequent Wittig-Horner coupling with triethyl phosphonoacetate using lithium chloride and diisopropylethylamine as condensing agents.¹⁵ The thus-obtained (E)alkene esters 7 were reduced with DIBALH to allylic alcohols 8, which were subjected to a Sharpless epoxidation¹⁶ using L-(+)-diethyl tartrate as chiral inductor to give S, S epoxy alcohols 9 in excellent chemical and optical yields as was evident from extensive GC analysis of 9 and product 10 derived thereof (the R, R, R diastereomers were only present in minor amounts, less than 5%).

After proper protection¹⁷ of the primary alcohol function, the triple bond was removed by hydrogenation together with the benzyl protecting group. In fact, this benzyl ether function was chosen as the protecting group in 3 for this purpose, allowing the introduction of an acetate at this position which is appropriate for the rest of the sequence. After liberation of the primary alcohol with fluoride. alcohols 13 were converted into diazo ketones 14 successively by oxidation with ruthenium oxide¹⁸ to the corresponding oxiranecarboxylic acids, treatment with isobutyl chloroformate to give mixed anhydrides, and addition of excess of diazomethane. It should be noted that an

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alternative to the ruthenium oxide method is a two-step procedure involving first a Swern oxidation to the aldehyde and then conversion to the epoxy acid using sodium chlorite.¹⁹ This last-mentioned methodology is more generally applicable. The yields of epoxy diazomethyl ketones based on epoxy alcohols 13 are in the range of 50-70% for both oxidation routes.

Irradiation of key intermediates 14 at 300 nm in ethanol resulted in γ -hydroxy α,β -unsaturated esters 15, which were immediately protected as silvl ethers (Scheme IV). It should be noted that for this protection a 2-fold excess of silvlating agent was required to achieve complete conversion. Chromatography of thus-obtained compounds 16 gave predominantly the E isomers, along with a small amount of the Z isomers. In all three cases the E:Z ratio amounted to 10:1. The formation of E as well as Z alkene esters has been explained previously.² In the present case, advantage was taken of this production of both geometrical isomers of alkene esters 16 as it enabled us to prepare the corresponding Z isomers of the target macrolides. Accordingly, both isomers of 16 were deprotected to hydroxy acids 17E and 17Z, respectively. This deprotection was performed with either lithium hydroxide in THF or sodium hydroxide in ethanol. The acetate function is readily saponified; however, the unsaturated ester was removed very slowly. The use of pig liver esterase was explored for the hydrolysis of the alkene ester; however, the results were disappointing.

For the macrolactonization of 17Eb, several methods were investigated, namely, the Masamune reaction using diethyl phosphochloridate,²⁰ closure with (trimethylsilyl)ethoxyethyne,²¹ and reaction with cyanuric chloride²² and

^{(12) 6-}Heptyn-1-ol was prepared in the following manner: tetrahydropyran and acetyl bromide were heated in the presence of ZnBr₂ (Ames, ; Islip, P. J. J. Chem. Soc. 1963, 4363). Removal of the acetyl function D.E with K₂CO₃ in MeOH gave 5-bromopentanol. Protection of the alcohol function with dihydropyran and coupling with lithium acetylide gave the

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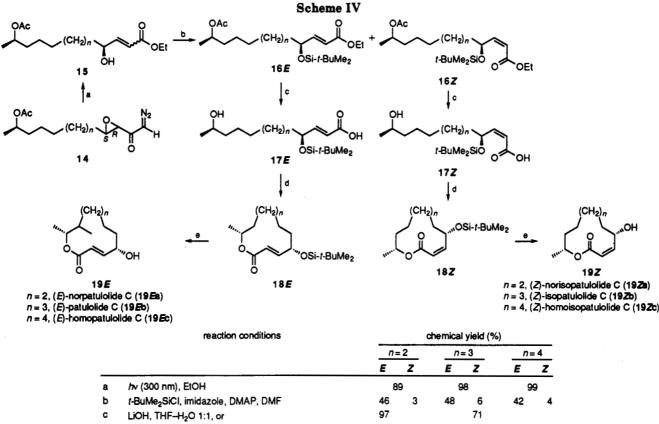
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NaOH, EtOH

d 2,6-Cl₂C₆H₃C(O)Cl, Et₃N, THF; DMAP, toluene

TBAF, THF, or

CH3COOH/THF/H2O 3:1:1

DMF/SOCl₂.²³ None of these conditions resulted in the desired lactonization. Better results were obtained by applying the Yamaguchi procedure,²⁴ as previously used by us in a slightly modified form.^{3,4,5,7} In all six cases, acceptable yields of silyl-protected macrolides 18 were obtained. However, during ring closure of 17Zb to 18Zb, considerable Z to E isomerization was observed, producing a mixture of 18Eb and 18Zb in a ratio of approximately 1:1. A similar isomerization has been noticed by Mori et al.²⁵ Apparently the ring strain in the macrolide causes isomerization probably induced by the (dimethylamino)pyridine (DMAP) via an addition/elimination reaction, although Z to E isomerization prior to the ring-closure reaction is a possibility that must be considered. Keck et $al.^{26}$ used a DMAP-catalyzed Z to E isomerization to obtain pure trans unsaturated thiol esters.

The final step involves the removal of the silyl protecting function, which was performed with TBAF in THF at 0 °C in the usual manner.¹⁷ However, these conditions failed for 19Zc, which had to be deprotected with 3:1:1 acetic

acid/THF/H₂O. All six products were purified by flash chromatography and characterized by spectral measurements [1H NMR (400 MHz), 13C NMR, IR, MS, and peak match].

97

28

66

99

21

49

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71

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64

The spectral features of 19Eb were identical with those obtained²⁷ from authentic natural patulolide C. This was not the case, however, for the optical rotation. For natural patulolide C, an $[\alpha]^{25}$ value of -1.89° [c 2, EtOH] has been reported,⁹ while for 19Eb, an $[\alpha]^{20}$ value of +6.6° [c 0.4, EtOH] was determined. X-ray diffraction analysis²⁸ of the p-bromobenzoate of 19Eb (mp 136-138 °C, lit.9 mp 134-135 °C) confirmed our conviction that the reported rotation for the natural product is not correct.

The detailed pattern of the ¹H NMR spectrum of (E)norpatulolide C (19Ea) was rather different from that of (E)-patulolide C (19Eb) (vide infra). Therefore, the structure of (E)-norpatulolide C (19Ea) was ascertained by X-ray diffraction analysis.²⁹ The absolute configuration was established for both compounds and is 4S,10R for (E)-norpatulolide C (19Ea) and 4S, 11R for (E)-patulolide C (19Eb).

It is of interest to note that (Z)-isopatulolide C has recently been isolated from the culture broth of P. urticae

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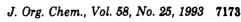
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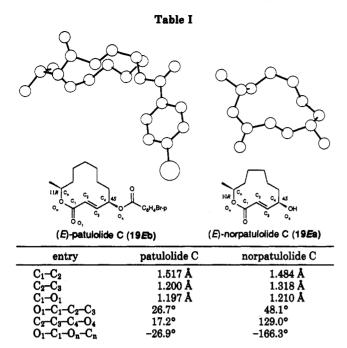
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mutant S11R59.³⁰ The configuration at C_{11} was assumed to be R as in all related patulolides; the R configuration was suggested for C_4 although no unambiguous evidence was provided. This natural compound has an $[\alpha]^{20}$ of -70.5° (c 2.4, EtOH),²⁷ which is almost the same value but opposite in sign of that observed for the synthetic material 19Zb ($[\alpha]^{20}$ _D +71°) described above. These data would suggest that the natural and synthetic compound are enantiomers, but their ¹H NMR spectra are distinctly different although both have the same absorption characteristics and are in full accord with the (Z)-isopatulolide structure. Our synthetic compound has the 4S,11Rconfiguration as is evident from its synthesis; therefore we suggest that the natural product mentioned above is diastereometric with 19Zb and has the 4R,11R configuration.

The conformations of the respective macrolides deserve some further comments. It is already apparent from the X-ray structures of (E)-norpatulolide C (19Ea) and (E)patulolide C (19Eb) that there is a considerable difference in spatial arrangement. In Table I the most important structural parameters taken from the X-ray structures are listed. The different torsion angles between the carbonyl and olefinic bonds are particularly noteworthy. These angles indicate that in both compounds ring strain enforces an out of plane twist of the enone moiety, most strikingly in the 11-membered ring lactone. As a consequence, there will be loss of conjugation in this enone unit. Nonplanarity of the enone moiety has also been observed for other macrocyclic α,β -unsaturated lactones.³¹

The conformational differences of these macrolides are also reflected in their spectral features. The essential spectral data of the six macrolides 19 are collected in Table II. Loss of conjugation in the alkene ester unit will result in a more isolated double bond character, hence, a less polarized double bond. Both in the ¹H NMR and ¹³C NMR such a diminished conjugation will lead to a smaller difference in chemical shift of the olefinic protons and carbon atoms, respectively, compared to the more conjugated case. In the UV spectrum, loss of conjugation results in a hypsochromic effect when compared with an unstrained "open" compound, for which ethyl (E)-4hydroxynon-2-enoate $(20)^2$ was taken as reference in the present case. The data in Table II reveal that in the Eseries (E)-norpatulolide C (19Ea) is the most strained molecule. The 12- and 13-membered ring lactones are much less deformed and show rather normal data compared with the open chain compound 20. The iso series (19Z) also exhibits considerable ring strain as is indicated by the difference in chemical shift of the olefinic protons and carbon atoms, respectively. The same is reflected in the UV spectra. Apparently both (Z)-norisopatulolide C and (Z)-homoisopatulolide C almost completely loose the alkene ester conjugation.

¹H NMR spectra of 19Ea have been recorded in solvents other than CDCl₃ and show a high solvent dependency. In particular, the positions of the olefinic protons in the spectrum change considerably. Figure 1 shows the chemical shifts of the olefinic protons when recorded in CDCl₃ and in CD₃CN. The spectra of 19Ea in acetone- d_6 and methanol- d_4 were very similar to that recorded in CD₃CN (Table III). In these polar solvents proton H_2 is observed at lower field than H₃. This can be interpreted in terms of a twist of the C=CC=O moiety, which causes deconjugation and a more localized charge on the carbonyl function. This twisted conformation is apparently stabilized by polar solvents. Another phenomenom could be the compressing effect of polar solvents on the apolar paraffinic methylene groups, causing a further increase of the torsion angle (Table I). The UV spectra of 19Ea are consistent with these NMR results. A maximum absorption at 205 nm was found when the spectrum was recorded in isooctane; in acetonitrile this absorption appeared at 194 nm. In general, for α,β -unsaturated carbonyl compounds³² the increase of solvent polarity will cause a red shift of the maximum absorption. In our case the opposite, a blue shift, was found. This solvent dependence is an interesting aspect in view of conformationally controlled reactions with macrolides, which appear frequently in the literature.33

In conclusion, the strategy for the total synthesis of patulolides as outlined in Scheme II successfully leads to the desired 11-, 12-, and 13-membered macrolides. In addition, the corresponding macrolides with a *cis* double bond could also be prepared. The six macrolides show remarkable differences in their conformational behavior.

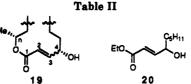
Experimental Section

General Procedures. ¹H NMR spectra were recorded on a Varian EM 390 (90 MHz, CW), a Bruker WH 90 (90 MHz, FT), a Bruker AM 100 (100 M Hz, FT), or a Bruker AM 400 (400 MHz, FT) spectrometer with TMS, CHCl₃, or CH₂Cl₂ as internal standard. IR spectra were recorded on a Perkin-Elmer 298 spectrophotometer and UV spectra on a Perkin-Elmer Lambda 5 UV/vis spectrophotometer. For mass spectroscopy a double focusing VG 7070 E was used. For the chemical ionization (CI) technique, methane was used as reacting gas. GC was performed

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	¹ H NMR (CDCl ₃ , ppm)								¹³ C NMR (CDCl ₃ , ppm)			UV (CH ₃ CN, nm)	
compd	δH_2	δH ₃	δH3-δH2	δH4	δHn	J_{23}	J ₃₄	J ₂₄	C1	C ₂	C ₈	λmax	Emax
19 <i>E</i> aª	6.34	6.46	0.12	4.40	4.65	16.1	8.9		168.87	122.61	145.70	194	8058
19 <i>E</i> b	6.07	6.82	0.75	4.45	5.04	16.5	6.0	1.2	168.09	121.49	149.69	208	7380
19 <i>E</i> c	6.02	6.89	0.87	4.50	5.11	15.8	6.9	1.0	166.93	121.46	149.38	206	10440
19Za	5.87	5.91	0.05	4.82	5.02	12.0	7.1		166.82	122.05	143.75	191	1529
19 <i>Z</i> b	5.79	6.02	0.23	5.12~5.02		11.9	8.5		166.38	121.08	146.80	201	8967
19Zc	5.88	5.90	0.04	5.26-5.13		11.4	8.4		166.78	121.93	146.21	<190	
20	6.02	6.93	0.91	4.28		16.0	5.0	1.5	167.2	119.30	151.30	208	8756

^a For ¹H-NMR spectrum in CD₃CN, see Experimental Section. ^b In isooctane, $\lambda_{max} = 205$ nm, $\epsilon_{max} = 7180$.

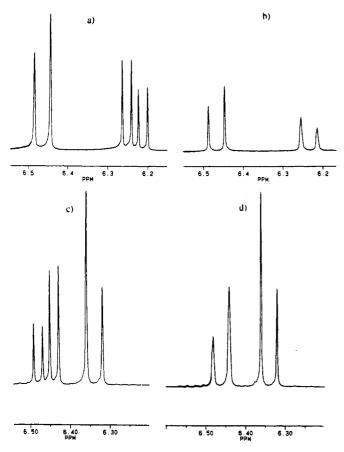


Figure 1. (a) ¹H-NMR spectrum of 19Ea in CD₃CN. (b) ¹H-NMR spectrum of 19Ea in CD₃CN, irradiated at H₄. (c) ¹H-NMR spectrum of 19Ea in CDCl₃. (d) ¹H-NMR spectrum of 19Ea in CDCl₃, irradiated at H₄.

Table III								
solvent	H_2 (ppm)	H_3 (ppm)	J ₂₃ (Hz)	J ₃₄ (Hz)				
CDCl _{3^a}	6.34	6.46	16.1	8.9				
CD ₃ CN ^a	6.46	6.23	16.2	9.2				
acetone-dsb	6.45	6.20	16.0	8.1				
CD3OD ⁶	6.59	6.29	16.0	8.4				

^a 400-MHz ¹H NMR. ^b 90-MHz ¹H NMR.

on a Hewlett-Packard 5790A or 5890 instrument equipped with a capillary HP cross-linked silicone $(25 \text{ m} \times 0.31 \text{ mm})$ column, connected to a HP 3390 or HP 5890 calculating integrator. Optical rotations were measured on a Perkin-Elmer 241 polarimeter. Melting points were determined on a Reichert Thermopan microscope and are uncorrected. Column chromatography was performed with silica gel 60H (Merck). A pressure of 1.5–2.0 bar was used to obtain the necessary flow rate. Dry solvents were obtained as follows. Petroleum ether (60-80 °C) was distilled from NaH. CH_2Cl_2 was distilled from P_2O_5 . Et_2O was predried on $CaCl_2$ and then distilled from CaH_2 . THF was distilled from LiAlH₄. Pyridine, Et_3N , diisopropylethylamine were distilled from KOH. DMSO was distilled from CaH₂. Toluene was distilled from sodium. All other solvents were of either p.a. or "reinst" quality. *n*-BuLi and DIBALH were used as stock solutions of 1.6 and 1.0 M in hexane, respectively.

(R)-8-(Tetrahydropyranyl-2-oxy)-4-octyn-2-ol (3a). n-BuLi (1.6 M in hexane, 56 mL, 90.3 mmol) was added to a solution of 2a (15.17 g, 90.3 mmol) in THF (200 mL) at -78 °C under an argon atmosphere. After 45 min at 0 °C, the reaction mixture was cooled to -50 °C and HMPA (25 mL) and (R)-methyloxirane (1) (5 g, 86 mmol) were rapidly added, and the reaction mixture was allowed to reach rt. After being stirred overnight, the dark brown reaction mixture was quenched with 50 mL of 5 N NH₄Cl and the layers were separated. The aqueous layer was extracted three times with hexane. The combined organic layers were washed with water, dried with MgSO4, filtered, and concentrated. Purification by flash chromatography (hexane/EtOAc 5:1) gave 9.46 g of 3a (41.9 mmol, 46%) and 3.38 g of recovered 2a. 3a: ¹H NMR (90 MHz, CDCl₃) δ 4.55 (s, 1H), 4.07–3.30 (m, 5H), 2.43-3.13 (m, 4H), 2.57 (s, 1H), 1.95-1.33 (m, 8H), 1.26 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3640-3100, 3580; MS (CI⁺) m/e (rel intensity) 227 $(M^+ + 1, 8)$, 209 (4), 143 (23), 125 (13), 85 (100).

(*R*)-9-(Tetrahydropyranyl-2-oxy)-4-nonyn-2-ol (3b). The procedure described for 3a was followed. From 12.7 g of 2b (70 mmol) and 4.4 g of (*R*)-methyloxirane (76 mmol), 11 g of 3b (45.8 mmol, 65%) was obtained after chromatography (hexane/EtOAc 3:1): ¹H NMR (90 MHz, CDCl₃) δ 4.57 (s, 1H), 4.20–3.25 (m, 5H), 2.43–2.05 (m, 4H), 2.00 (s, 1H), 1.92–1.37 (m, 10H), 1.23 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3640–3200; MS (CI⁺) m/e (rel intensity) 241 (M⁺ + 1, 3), 223 (7), 205 (1), 157 (62), 139 (53), 85 (100).

(*R*)-10-(Tetrahydropyranyl-2-oxy)-4-decyn-2-ol (3c). The procedure described for 3a was followed. From 13.5 g of 2c (69 mmol) and 4.0 g of (*R*)-methyloxirane (69 mmol), 15.0 g of 3c (59 mmol, 86%) was obtained after chromatography (hexane/EtOAc 3:1): ¹H NMR (90 MHz, CDCl₃) δ 4.59 (bs, 1H), 4.13–4.20 (m, 5H), 2.43–2.00 (m, 4H), 1.93–1.33 (m, 12H), 1.23 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3620–3120, 3580; MS (CI⁺) m/e (rel intensity) 255 (M⁺ + 1, 15), 237 (2), 171 (85), 153 (9), 85 (100).

(R)-7-(Benzyloxy)-1-(tetrahydropyranyl-2-oxy)-4-octyne (4a). A solution of 3a (9.3 g, 41 mmol) in 70 mL of THF under an argon atmosphere was treated with NaH (1.34 g, 44.5 mmol), followed by addition of *n*-Bu₄NI (151 mg, 0.41 mmol) and benzyl bromide (7 g, 41 mmol). After being stirred overnight, the reaction mixture was quenched with 70 mL of water and THF was evaporated. The resulting mixture was extracted three times with ether. The organic layers were combined, washed with water, dried over MgSO₄, and concentrated *in vacuo*. Chromatography (hexane/EtOAc 10:1) gave 4a (9.69 g, 30.7 mmol) in a yield of 75%: ¹H NMR (90 MHz, CDCl₃) δ 7.40–7.12 (m, 5H), 4.57 (s, 3H), 4.04–3.31 (m, 5H), 2.53–2.09 (m, 4H), 2.00–1.40 (m, 8H), 1.29 (d, 3H, J = 6 Hz); MS (CI⁺) m/e (rel intensity) 317 (M⁺ + 1, 2), 233 (32), 215 (15), 91 (67), 85 (100). (*R*)-8-(Benzyloxy)-1-(tetrahydropyranyl-2-oxy)-5-nonyne (4b). The procedure described for 4a was followed. From 11.36 g of 3b (47.3 mmol), 10.6 g of 4b (32.1 mmol) was obtained after chromatography (hexane/EtOAc 7:1) in a yield of 68%: ¹H NMR (90 MHz, CDCl₃) δ 7.50–7.20 (m, 5H), 4.50 (s, 3H), 4.10– 3.22 (m, 5H), 2.50–2.05 (m, 4H), 1.90–1.40 (m, 10H), 1.27 (d, 3H, J = 6 Hz); MS (CI⁺) m/e (rel intensity) 331 (M⁺ + 1, 19), 247 (100), 229 (22).

(R)-9-(Benzyloxy)-1-(tetrahydropyranyl-2-oxy)-6-decyne (4c). The procedure described for 4a was followed. From 14.55 g of 3c (57.3 mmol), 19 g of crude 4c was obtained as a slightly yellowish oil which was sufficiently pure for further use: ¹H NMR (90 MHz, CDCl₃) δ 7.57-7.12 (m, 5H), 4.77-4.55 (m, 3H), 4.07-3.20 (m, 5H), 2.67-2.00 (m, 4H), 2.00-1.20 (m, 12H), 1.27 (d, 3H, J = 6 Hz).

(*R*)-7-(Benzyloxy)-4-octyn-1-ol (5a). p-TsOH (160 mg, 0.93 mmol) was added to a solution of 4a (9.90 g, 31 mmol) in 70 mL of MeOH. After the solution was stirred for 2 h, 4 mL of saturated NaHCO₃ was added and the MeOH was evaporated. Water was added and the solution was extracted three times with ether. The combined organic layers were dried over MgSO₄ and concentrated in vacuo. 5a (6.67 g) was obtained as a colorless oil, after chromatography (hexane/EtOAc 3:1), in a yield of 93%: ¹H NMR (90 MHz, CDCl₃) δ 7.50-7.10 (m, 5H), 4.53 (s, 2H), 3.83-3.48 (m, 3H), 2.48-2.10 (m, 5H), 1.70 (sext., 2H), 1.23 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3630, 3700-3200; MS (CI⁺) m/e (rel intensity) 233 (M⁺ + 1, 49), 215 (100), 187 (14), 125 (22), 91 (90).

(*R*)-8-(Benzyloxy)-5-nonyn-1-ol (5b). The procedure described for 5a was followed. From 10.3 g of 4b (31.2 mmol), 7.6 g of 5b was obtained which was sufficiently pure (98% on GC) for further use: ¹H NMR (90 MHz, CDCl₃) δ 7.47-7.10 (m, 5H), 4.55 (s, 2H), 3.80-3.43 (m, 3H), 2.50-2.07 (m, 4H), 1.98 (bs, 1H), 1.83-1.43 (m, 4H), 1.28 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3640, 3600-3100; MS (CI⁺) m/e (rel intensity) 247 (M⁺ + 1, 3), 229 (7), 211 (4), 173 (25), 135 (32), 105 (71), 91 (90).

(*R*)-9-(Benzyloxy)-6-decyn-1-ol (5c). The procedure described for 5a was used. From 19 g of crude 4c (55 mmol), 13.44 g of pure 5c (51.7 mmol, 94%) was obtained, after chromatography (hexane/EtOAc 2:1): ¹H NMR (90 MHz, CDCl₃) δ 7.43–7.13 (m, 5H), 4.53 (s, 2H), 3.83–3.43 (m, 3H), 2.70–1.90 (m, 4H), 1.80 (bs, 1H), 1.70–1.37 (m, 6H), 1.23 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3630, 3600–3100; MS (CI⁺) m/e (rel intensity) 261 (M⁺ + 1, 2), 243 (21), 225 (1), 91 (100).

(R)-7-(Benzyloxy)-4-octynal (6a). Oxalyl chloride (2.49 mL, 28.7 mmol) was dissolved in 60 mL of CH₂Cl₂ at -78 °C under an argon atmosphere and DMSO (4.29 mL, 60.6 mmol) in 8 mL of CH2Cl2 and 5a (6.66 g, 28.7 mmol) in CH2Cl2 (20 mL) was added. After 20 min of stirring at -78 °C, Et₃N (20 mL) was added and the reaction mixture was allowed to warm up to rt. Water was added and then the solution was extracted three times with CH₂Cl₂. The combined organic layers were washed with saturated NaCl, 1% HCl (twice), and 5% Na₂CO₃ (twice). The organic layer was dried over MgSO4 and concentrated in vacuo, giving aldehyde 6a (6.18 g, 26.9 mmol) almost quantitatively. Because of the low stability of aldehydes in general, products 6 were not purified but directly used in the next step. IR showed that no starting material was present. ¹H NMR (90 MHz, CDCl₃): δ 9.77 (s, 1H), 7.40-7.10 (m, 5H), 4.52 (s, 2H), 3.62 (sext., 1H), 2.70–2.13 (m, 6H), 1.25 (d, 3H, J = 6 Hz). IR (CCl₄, cm⁻¹): 1725

(R)-8-(Benzyloxy)-5-nonynal (6b). The procedure described for 6a was used. From 11.0 g of 5b (44.7 mmol), 10.9 g of crude 6b was obtained, which was sufficiently pure for further use: ¹H NMR (90 MHz, CDCl₃) δ 9.73 (s, 1H), 7.50–7.13 (m, 5H), 4.53 (s, 2H), 3.63 (sext., 1H), 2.67–2.07 (m, 6H), 1.97–1.58 (m, 2H), 1.27 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 1725.

(R)-9-(Benzyloxy)-6-decynal (6c). The procedure described for 6a was used. From 5c (12.44 g), 6c was obtained in an almost quantitative yield: IR (CCl₄, cm⁻¹) 1725.

Ethyl (2E,9R)-9-(Benzyloxy)dec-2-en-6-ynoate (7a). LiCl (1.36g) was suspended in CH₃CN under argon. To this suspension were added triethyl phosphonoacetate (7.6 g, 32 mmol) and diisopropylethylamine (3.6 g, 28 mmol). After 15 min of stirring at rt, 6a (6.18 g, 26.9 mmol) in CH₃CN (20 mL) was added. The reaction was monitored by GC. After the solution was stirred

overnight, the solvent was concentrated under reduced pressure and water was added. This mixture was extracted three times with ether. The organic layers were dried over MgSO₄ and concentrated *in vacuo*. After chromatography (hexane/EtOAc 10:1), 6:00 g of 7a (20 mmol) was obtained in 74% yield: ¹H NMR (90 MHz, CDCl₃) δ 7.58–7.18 (m, 5H), 7.18–6.77 (m, 1H), 5:88 (d, 1H, J = 16:0 Hz), 4:56 (s, 2H), 4:20 (q, 2H, J = 7.3 Hz), 3:66 (sext., 1H), 2:70–2:11 (m, 6H), 1:29 (d, 3H, J = 6.0 Hz), 1:27 (t, 3H, J = 7.0 Hz); IR (CCl₄, cm⁻¹) 1715, 1650; MS (CI⁺) m/e (rel intensity) 301 (M⁺ + 1, 3), 283 (1), 269 (10), 255 (3), 209 (11), 193 (11), 91 (100).

Ethyl (2E,10R)-10-(Benzyloxy)undec-2-en-7-ynoate (7b). The procedure described for 7a was followed. From crude 6b (10.9 g) and triethyl phosphonoacetate (10.5 mL, 52.8 mmol), 7b (10.39 g, 33.0 mmol, 75%) was obtained after chromatography (hexane/EtOAc 10:1): ¹H NMR (90 MHz, CDCl₃) δ 7.50–7.25 (m, 5H), 6.97 (dt, 1H, J = 15.0, 6.8 Hz), 5.85 (d, 1H, J = 15.0 Hz), 4.58 (s, 2H), 4.17 (q, 2H, J = 7.5 Hz), 3.67 (sext., 1H), 2.60–2.05 (m, 6H), 1.90–1.47 (m, 2H), 1.43–1.13 (m, 6H); IR (CCl₄, cm⁻¹) 1720, 1655; MS (CI⁺) m/e (rel intensity) 315 (M⁺ + 1, 100), 297 (11), 269 (13), 223 (22), 207 (13), 91 (82).

Ethyl (2E,11R)-11-(Benzyloxy)dodec-2-en-8-ynoate (7c). The procedure described for 7a was followed. From 6c (12.3 g) and triethyl phosphonoacetate (12.86 g, 57.4 mmol) was obtained, after chromatography (hexane/EtOAc 10:1), 7c (13.66 g, 41.6 mmol, 87%): ¹H NMR (90 MHz, CDCl₃) δ 7.47-7.18 (m, 5H), 6.93 (dt, 1H, J = 15.0, 6.8 Hz), 5.83 (d, 1H, J = 15.0 Hz), 458 (s, 2H), 4.17 (q, 2H, J = 7.0 Hz), 3.65 (sext., 1H), 2.70-1.97 (m, 6H), 1.77-1.42 (m, 4H), 1.42-1.20 (m, 6H); IR (CCl₄, cm⁻¹) 1720, 1655; MS (CI⁺) m/e (rel intensity) 329 (M⁺ + 1, 25), 311 (3), 283 (4), 265 (4), 237 (9), 91 (100).

(2E,9R)-9-(Benzyloxy)dec-2-en-6-yn-1-ol (8a). DIBALH (41 mL) was added to a solution of 7a (6.0 g, 20 mmol) in ether (120 mL) at 0 °C under argon. After 2.5 h the reaction was complete. The reaction mixture was quenched with Na₂- $SO_4 \cdot 10H_2O(7g)$. This mixture was stirred for 1 h. After filtration of the insoluble salts, the filtrate was dried over MgSO4 and concentrated in vacuo, giving 6.1 g of crude product. GC analysis showed that still 35% of starting material was present; separation by chromatography (hexane/EtOAc 3:1) gave pure 8a (3.34 g, 12.9 mmol) and starting material 7a (1.06 g, 3.5 mmol) which was again treated with DIBALH to give, after work up and chromatography, 0.65 g of 8a (total yield 78%): ¹H NMR (90 MHz, CDCl₃) § 7.50-7.20 (m, 5H), 5.77-5.60 (m, 2H), 4.53 (m, 2H), 4.00 (d, 2H, J = 3 Hz), 3.62 (sext., 1H), 2.47-2.13 (m, 6H), 2.00 (bs,1H), 1.25 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3610, 3700–3200; MS $(CI^+) m/e$ (rel intensity) 259 $(M^+ + 1, 5)$, 241 (14), 223 (11), 197 (25), 183 (40), 135 (58), 91 (100).

(2E,10R)-10-(Benzyloxy)undec-2-en-7-yn-1-ol (8b). The procedure described for 8a was followed. From 7b (10.0 g, 32 mmol), 5.00 g of 8b (18.4 mmol, 57%) was obtained, after chromatography (hexane/EtOAc 3:1). Repetition of the procedure with fresh DIBALH gave 8b in 80% yield: ¹H NMR (90 MHz, CDCl₃) δ 7.50–7.10 (m, 5H), 5.77–5.60 (m, 2H), 4.57 (m, 2H), 4.00 (d, 2H, J = 3 Hz), 3.63 (sext., 1H), 2.60–2.00 (m, 7H), 1.83–1.40 (m, 2H), 1.27 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3620, 3600–3250, 1670.

(2E,11*R*)-11-(Benzyloxy)undec-2-en-8-yn-1-ol (8c). The procedure described for 8a was followed. From 7c (12.66 g, 38.6 mmol) and fresh DIBALH (85 mL), 10.73 g of 8c (37.5 mmol, 97%) was obtained, which was sufficiently pure (98% on GC) for further use: ¹H NMR (90 MHz, CDCl₃) δ 7.53–7.13 (m, 5H, 5.76–5.55 (m, 2H), 4.57 (m, 2H), 4.03 (d, 2H, J = 3 Hz), 3.83–3.40 (m, 1H), 2.67–1.80 (m, 7H), 1.73–1.17 (m, 4H), 1.28 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3620, 3700–3100; MS (CI⁺) m/e (rel intensity) 287 (M⁺ + 1, 1), 269 (5), 199 (8), 173 (25), 135 (45), 91 (100).

(2S,3S,9R)-9-(Benzyloxy)-2,3-epoxy-6-decyn-1-ol (9a). (+)-Diethyl L-tartrate [(+)-L-DET] (113 μ L, 0.66 mmol) and Ti(O*i*-Pr)₄ (164 μ L, 0.55 mmol) were added to a suspension of 1 g of powdered molecular sieves (4 Å) in 100 mL of CH₂Cl₂ at -20 °C under argon. Then t-BuOOH (5.4 mL, 22 mmol, M = 4.08 in dichloroethane) was slowly introduced. After being stirred between -20 and -15 °C for 0.5 h, a solution of 8a (2.83 g, 11 mmol) in CH₂Cl₂ (5 mL) was added in 20 min. The reaction was monitored by TLC. After 3.5 h of stirring at -20 °C to -15 °C, the reaction mixture was quenched with a precooled solution of FeSO₄ (3.7 g) and DL-tartaric acid (1.1 g) in water (11 mL). After the solution was stirred for 5 min, the layers were separated, the aqueous layer was extracted twice with ether, and the combined organic layers were cooled to 0 °C and then treated with a precooled 30% NaOH (3 mL) in saturated NaCl solution. This mixture was stirred for another 1 h. Water was added and the solution was extracted three times with ether. The combined organic layers were dried and concentrated *in vacuo*. After chromatography (hexane/EtOAc 1:1), 2.83 g of pure **9a** (10.3 mmol, 94%) was obtained as a colorless oil: ¹H NMR (90 MHz, CDCl₃) δ 7.53-7.17 (m, 5H), 4.53 (s, 2H), 3.93-3.33 (m, 3H), 3.12-2.83 (m, 2H), 2.62-2.07 (m, 5H), 1.93-1.53 (m, 2H), 1.23 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3600, 3650-3200; MS (Cl⁺) m/e (rel intensity) 275 (M⁺ + 1, 21), 257 (40), 239 (8), 231 (51), 167 (34), 91 (100); $[\alpha]^{20}_{D} - 17.7^{\circ}$ (c 0.94, CHCl₃).

(2S,3S,10R)-10-(Benzyloxy)-2,3-epoxy-7-undecyn-1-ol (9b). The procedure described for 9a was followed. From 8b (5.00 g, 18.4 mmol), 4.86 g of 9b was obtained, after chromatography (hexane/EtOAc 2:1), in a yield of 92%: ¹H NMR (90 MHz, CDCl₃) δ 7.50–7.20 (m, 5H), 4.56 (s, 2H), 4.00–3.33 (m, 3H), 3.07–2.80 (m, 2H), 2.67–2.00 (m, 5H), 1.90–1.43 (m, 4H), 1.27 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3650–3200; MS (CI⁺) m/e (rel intensity) 287 (M⁺ – 1, 0.4), 271 (0.14), 181 (4), 91 (52).

(2S,3S,11R)-11-(Benzyloxy)-2,3-epoxy-8-dodecyn-1-ol (9c). The procedure described for 9a was followed. From 8c (10.73 g, 38.6 mmol), 9.13 g of 9c (30.2 mmol, 78%) was obtained, after chromatography (petroleum ether (60-80 °C)/EtOAc 2:1); ¹H NMR (90 MHz, CDCl₃) δ 7.50-7.13 (m, 5H), 4.53 (s, 2H), 3.97-3.30 (m, 3H), 3.00-2.73 (m, 2H), 2.67-1.93 (m, 5H), 1.83-1.20 (m, 6H), 1.27 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3650-3200; MS (CI⁺) m/e (rel intensity) 301 (M⁺ - 1, 0.4), 285 (5), 173 (7), 105 (68), 91 (100); $[\alpha]^{20}_D$ -11.9° (c 0.96, CHCl₃).

(2R,8S,9S)-2-(Benzyloxy)-8,9-epoxy-10-(tert-butyldimethylsiloxy)-4-decyne (10a). Imidazole (1.85 g, 27.2 mmol) and TBDMSCl (1.75 g, 11.6 mmol), dissolved in DMF (36 mL), were added to a solution of 9a (3 g, 10.9 mmol) in DMF (36 mL) under argon. The reaction was monitored by TLC. After 2 h of stirring, water was added and the mixture was extracted three times with hexane. The combined organic layers were washed with water, dried (MgSO₄), and concentrated *in vacuo*, giving 3.97 g of crude 10a (10.2 mmol, 94%), which was sufficiently pure for further use: ¹H NMR (90 MHz, CDCl₃) δ 7.45–7.18 (m, 5H, Ph), 4.60 (s, 2H), 4.02–3.57 (m, 3H), 3.13–2.88 (m, 2H), 2.60–2.10 (m, 4H), 2.00–1.67 (m, 2H), 1.37 (d, 3H, J = 6 Hz), 1.03 (s, 9H), 0.18 (s, 6H); IR (CCL, cm⁻¹) 1250; MS (CI⁺) m/e (rel intensity) 389 (M⁺ + 1, 2), 371 (12), 281 (17), 257 (8), 195 (28), 149 (17), 91 (100).

(2R,9S,10S)-2-(Benzyloxy)-9,10-epoxy-11-(*tert*-butyldimethylsiloxy)-4-undecyne (10b). The procedure described for 10a was followed. From 9b (4.86 g, 16.9 mmol), 6.74 g of 10b (16.8 mmol, 99%) was obtained after chromatography (hexane/ EtOAc 10:1): ¹H NMR (90 MHz, CDCl₃) δ 7.56–7.20 (m, 5H), 4.60 (s, 2H), 3.97–3.37 (m, 3H), 3.10–2.77 (m, 2H), 2.57–2.10 (m, 4H), 1.97–1.53 (m, 2H), 1.37 (d, 3H, J = 6 Hz), 0.98 (s, 9H), 0.18 (s, 6H); IR (CCl₄, cm⁻¹) 1250.

(2R,10S,11S)-2-(Benzyloxy)-10,11-epoxy-12-(*tert*-butyldimethylsiloxy)-4-dodecyne (10c). The procedure described for 10a was followed. From 9c (7.62 g, 25.2 mmol), 10.55 g of crude 10c (25 mmol, 99%) was obtained which was sufficiently pure (99% on GC) for further use: ¹H NMR (90 MHz, CDCl₃) δ 7.60-7.25 (m, 5H), 4.67 (s, 2H), 4.00-3.53 (m, 3H), 3.00-2.80 (m, 2H), 2.60-2.10 (m, 4H), 1.97-1.50 (m, 2H), 1.40 (d, 3H, J = 6 Hz), 1.03 (s, 9H), 0.23 (s, 6H); IR (CCl₄, cm⁻¹) 1250; MS (CI⁺) m/e (rel intensity) 417 (M⁺ + 1, 0.5), 399 (1.4), 309 (4), 267 (4), 117 (36), 91 (100).

(2R,8S,9S)-8,9-Epoxy-10-(*tert*-butyldimethylsiloxy)decan-2-ol (11a). 10a (3.83 g, 9.86 mmol) was dissolved in absolute EtOH (200 mL). This solution was hydrogenated for 2.5 h with \approx 100 mg of Pd/C (10%) as a catalyst. The catalyst was removed by filtration and the filtrate was concentrated *in vacuo*. After chromatography (hexane/EtOAc 3:1), pure 11a (2.14 g, 7.1 mmol) was obtained in 72% yield: ¹H NMR (90 MHz, CDCl₃) δ 3.93-3.50 (m, 3H), 2.92-2.70 (m, 2H), 1.93-1.30 (m, 11H), 1.30 (d, 3H, J = 6 Hz), 0.93 (s, 9H), 0.12 (s, 6H); IR (CCL, cm⁻¹) 3620, 3700-3140, 1250; MS (CI⁺) m/e (rel intensity) 303 (M⁺ + 1, 10), 285 (44), 267 (13), 227 (18), 171 (17), 153 (67), 135 (100), 109 (68). (2R,9S,10S)-9,10-Epoxy-11-(*tert*-butyldimethylsiloxy)undecan-2-ol (11b). The procedure described for 11a was followed. Conversion of 10b (5.27 g, 13.1 mmol) gave 2.70 g of 11b after chromatography (hexane/EtOAc 4:1): ¹H NMR (90 MHz, CDCl₃) δ 3.95-3.50 (m, 3H), 2.93-2.67 (m, 2H), 1.80-1.25 (m, 13H), 1.30 (d, 3H, J = 6.3 Hz), 0.92 s, 9H), 0.10 (s, 6H); IR (CCl₄, cm⁻¹) 3620, 3600-3200, 1250; MS (CI⁺) m/e (rel intensity) 317 (M⁺ + 1, 8), 299 (57), 281 (10), 241 (26), 149 (100), 117 (98).

(2R,10S,11S)-10,11-Epoxy-12-(*tert*-butyldimethylsiloxy)dodecan-2-ol (11c). The procedure described for 11a was followed. 10c (4.85 g) gave 11c (2.70 g, 8.2 mmol) in a yield of 70% after chromatography (petroleum ether (60-80 °C)/EtOAc 4:1): ¹H NMR (90 MHz, CDCl₃) δ 4.00-3.53 (m, 3H), 3.00-2.73 (m, 2H), 1.80-1.20 (m, 15H), 1.27 (d, 3H, J = 7.0 Hz), 0.97 (s, 9H), 0.20 (s, 6H); IR (CCl₄, cm⁻¹) 3620, 3650-3140; MS (CI⁺) m/e (rel intensity) 331 (M⁺ + 1, 49), 313 (100), 295 (21), 163 (100).

(2R,8S,9S)-2-Acetoxy-8,9-epoxy-10-(tert-butydimethylsiloxy)decane (12a). Ac₂O (2.5 g, 24.5 mmol) and DMAP (10 mg) were added to a solution of 11a (2.04 g, 6.74 mmol) in pyridine (17 mL). After the solution was stirred for 1.5 h, the reaction was complete and ice was added. This mixture was extracted three times with ether. The organic layers were washed with saturated NaHCO₃ and water, dried (MgSO₄), and concentrated *in vacuo*. After stripping with toluene, 2.32 g of 12a (6.74 mmol, 100%) was obtained: ¹H NMR (90 MHz, CDCl₃) δ 5.07-4.60 (m, 1H), 3.90-3.47 (m, 2H), 2.95-2.63 (m, 2H), 2.00 (s, 3H), 1.77-1.27 (m, 10H), 1.17 (d, 3H, J = 6 Hz), 0.90 (s, 9H), 0.10 (s, 6H); IR (CCl₄, cm⁻¹) 1730, 1250.

(2R,9S,10S)-2-Acetoxy-9,10-epoxy-11-(*tert*-butyldimethylsiloxy)undecane (12b). The procedure described for 12a was followed. From 11b (2.71 g, 8.6 mmol), 2.84 g of 12b (7.93 mmol, 92%) was obtained after the usual workup. This compound was sufficiently pure (90% on GC) for further use: ¹H NMR (90 MHz, CDCl₃) δ 4.93 (sext., 1H), 3.95–3.55 (m, 2H), 2.98–2.75 (m, 2H), 2.07 (s, 3H), 1.83–1.20 (m, 12H), 1.17 (d, 3H, J = 6 Hz), 1.00 (s, 9H), 0.17 (s, 6H); IR (CCl₄, cm⁻¹) 1730, 1240.

(2R,10S,11S)-2-Acetoxy-10,11-epoxy-12-(*tert*-butyldimethylsiloxy)dodecane (12c). The procedure described for 12a was followed. From 11c (5.6 g, 17 mmol), 6.09 g of 12c (16.4 mmol, 96%) was obtained after workup, which was sufficiently pure (99% on GC) for further use: ¹H NMR (90 MHz, CDCl₃) δ 4.90 (sext., 1H), 3.93-3.53 (m, 2H), 2.97-2.67 (m, 2H), 2.00 (s, 3H), 2.73-2.17 (m, 12H), 2.22 (d, 3H, J = 6 Hz), 0.93 (s, 9H), 0.10 (s, 6H); IR (CCl₄, cm⁻¹) 1740, 1240; MS (CI⁺) m/e (rel intensity) 373 (M⁺ + 1, 26), 355 (48), 313 (21), 295 (15), 255 (36), 163 (100).

(2S,3S,9R)-9-Acetoxy-2,3-epoxydecan-1-ol (13a). n-Bu₄NF (6.8 mL, 1 M in THF) was added to a solution of 12a (3.34 g, 6.8 mmol) in 40 mL of THF at 0 °C via syringe. The reaction was monitored by GC. After 15 min, the reaction was complete and saturated NH₄Cl was added. This mixture was extracted three times with ether. The combined organic layers were dried over MgSO₄ and concentrated *in vacuo*. Chromatography (hexane/ EtOAc 1:1) gave 13a (1.41 g, 6.12 mmol) in 90% yield: ¹H NMR (90 MHz, CDCl₃) δ 4.90 (sext., 1H), 4.00–3.40 (m, 2H), 3.03–2.73 (m, 3H), 2.00 (s, 3H), 1.77–1.27 (m, 10H), 1.20 (d, 3H, J = 6 Hz); IR (CCL₄, cm⁻¹) 3650–3200, 1725; MS (CI⁺) m/e (rel intensity) 231 (M⁺ + 1, 1), 213 (1), 171 (13), 153 (21), 135 (90), 109 (100).

(2S,3S,10R)-10-Acetoxy-2,3-epoxyundecan-1-ol (13b). The procedure described for 13a was followed. From 12b (2.84 g, 7.93 mmol), 13b (1.77 g, 7.25 mmol) was obtained in 92% yield after chromatography (hexane/EtOAc 1:1): ¹H NMR (90 MHz, CDCl₃) δ 4.66 (sext., 1H), 3.85 and 3.61 (dq, 2H, J = 12, 2 Hz, respectively, J = 12, 4 Hz), 3.05–2.80 (m, 2H), 2.40 (bs, 1H), 2.03 (s, 3H), 1.75–1.10 (m, 12H), 1.17 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3600–3200, 1730.

(2S,3S,11R)-11-Acetoxy-2,3-epoxydodecan-1-ol (13c). The procedure described for 13a was followed. From 12c (6.0 g, 16.1 mmol), 4.15 g of crude 13c (16 mmol, 99%) was formed: ¹H NMR (90 MHz, CDCl₃) δ 5.10-4.67 (m, 1H), 4.05-3.40 (m, 2H), 3.07-2.73 (m, 2H), 2.50 (bs, 1H), 2.00 (s, 3H), 1.73-1.10 (m, 14H), 1.20 (d, 3H, J = 6 Hz); IR (CCl₄, cm⁻¹) 3650-3200, 1730; MS (CI⁺) m/e (rel intensity) 259 (M⁺ + 1, 42), 241 (10), 199 (70), 181 (32), 163 (100).

(3R,4S,10R)-10-Acetoxy-1-diazo-3,4-epoxyundecan-2one (14a). NaIO₄ (4.2 g, 19.6 mmol, 3.1 molar equiv) and RuCl₃·H₂O (31 mg, 2.2 mol %) were added to a solution of 13a

(1.45 g, 6.32 mmol) in CCl₄ (13 mL), CH₃CN (13 mL), and H₂O (20 mL). The reaction was monitored by TLC. After the solution was stirred for 2 h, CH₂Cl₂ (30 mL) and water (30 mL) were added and the layers were separated. The aqueous layer was extracted three times with CH2Cl2. The organic layer was dried over MgSO4 and concentrated in vacuo. The black residue was dissolved in dry ether (40 mL) and cooled to 0 °C under argon atmosphere. Isobutyl chloroformate (860 mg, 6.32 mmol) and Et₃N (960 mg, 9.48 mmol) were added to this solution. This mixture was stirred for 1 h at 0 °C. The precipitate was filtered off under argon flow and the filtrate was added to an excess of a diazomethane solution $(100 \text{ mL}, \approx 0.3 \text{ M})$ in ether. This mixture was left overnight. Excess diazomethane was removed by flushing with nitrogen. After evaporation, the residue was purified by chromatography (hexane/EtOAc 3:1), giving 14a (1.07g, 4.0 mmol) in 63% yield (yellow oil): 1H NMR (90 MHz, CDCl₃) & 5.48 (s, 1H), 4.87 (sext., 1H), 3.20 (d, 1H, J = 2 Hz), 3.07–2.87 (m, 1H), 2.00 (m, 3H), 1.73–1.25 (m, 10H), 1.17 (d, 3H, J = 7.0 Hz); IR (CCl₄, cm⁻¹) 3120, 2120, 1725, 1640; MS (CI⁺) m/e (rel intensity) $269 (M^+ + 1, 100), 241 (9), 181 (17), 163 (7), 135 (20), 121 (30),$ 109 (30), 95 (49), 81 (48).

(3R,4S,11R)-11-Acetoxy-1-diazo-3,4-epoxydodecan-2-one (14b). The procedure described for 14a was followed. From 13b (1.77 g, 7.25 mmol), 1.29 g of 14b (4.57 mmol, 63%) was obtained as a yellow oil after chromatography (hexane/EtOAc 3:1): ¹H NMR (90 MHz, CDCl₃) δ 5.47 (s, 1H), 4.88 (sext., 1H), 3.20 (d, 1H, J = 1.5 Hz), 3.07–2.83 (m, 1H), 2.02 (m, 3H), 1.77– 1.20 (m, 12H), 1.10 (d, 3H, J = 6.0 Hz); IR (CCl₄, cm⁻¹) 3120, 2120, 1725, 1640; MS (CI⁺) m/e (rel intensity) 283 (M⁺ + 1, 31), 255 (4), 223 (17), 195 (9), 177 (6), 135 (22), 109 (26), 95 (40), 81 (57), 69 (43), 55 (68), 43 (100).

(3R,4S,12R)-12-Acetoxy-1-diazo-3,4-epoxytridecan-2one (14c). The procedure described for 14a was followed. From 13c (3.66 g, 14.2 mmol), 2.90 g of 14c (9.80 mmol, 69%) was obtained as a yellow oil after chromatography (petroleum ether/ EtOAc 3:1): ¹H NMR (90 MHz, CDCl₃) δ 5.47 (s, 1H), 4.90 (sext., 1H), 3.17 (d, 1H, J = 2.0 Hz), 3.07–2.83 (m, 1H), 2.00 (m, 3H), 1.87–1.33 (m, 14H), 1.20 (d, 3H, J = 6.0 Hz); IR (CCl₄, cm⁻¹) 3120, 2120, 1730, 1640; MS (CI⁺) m/e (rel intensity) 297 (M⁺ + 1, 18), 269 (5), 237 (12), 209 (7), 191 (4), 149 (16), 111 (18), 95 (35), 81 (41), 69 (33), 55 (39).

Ethyl (2E/Z,4S,10R)-10-Acetoxy-4-hydroxy-2-undecenoate (15a). A nitrogen-flushed solution of 14a (1.07 g, 3.98 mmol) in EtOH (500 mL) was irradiated with UV light (300 nm). The rearrangement was monitored by IR spectroscopy (disappearance of the diazo absorption at 2120 cm⁻¹) and was complete after 2 h. Evaporation of the EtOH gave product 15a as a light brown oil (1.01 g, 89%) which was immediately used in the next step.

Ethyl (2E/Z,4S,11R)-11-Acetoxy-4-hydroxy-2-dodecenoate (15b). The procedure described for 15a was followed. Conversion of 1.25 g of 14b (4.43 mmol) gave 1.30 g of crude 15b (98%) as a brown/yellow oil which was immediately used in the next step.

Ethyl (2E/Z,4S,12R)-12-Acetoxy-4-hydroxy-2-tridecenoate (15c). The procedure described for 15a was followed. From 14c (2.58 g, 8.72 mmol), crude 15c (2.7 g, 99%) was obtained, which was immediately used in the next step.

Ethyl (2E/Z,4S,10R)-10-Acetoxy-4-(tert-butyldimethylsiloxy)-2-undecenoate (16Ea) and 16Za). Imidazole (600 mg, 8.85 mmol) and TBDMSCl (1.07 g, 7.08 mmol) in DMF (10 mL) were added to a stirred solution of crude 15a (1.01 g, 3.54 mmol) in DMF (15 mL) under argon. Finally a catalytic amount of DMAP was added. After stirring for 2.5 h, the reaction was quenched with 10 mL of water. This mixture was extracted three times with hexane. The combined organic layers were washed with water, dried (MgSO₄), and concentrated in vacuo. The residue was purified by chromatography (hexane/EtOAc 15:1 later 5:1), giving pure 16Ea (656 mg, 1.64 mmol, 46%) and 16Za (48 mg, 0.12 mmol, 3.4%). 16Ea: ¹H NMR (90 MHz, CDCl₃) δ $6.70 \,(dd, 1H, J = 16.0, 7.0 \,Hz), 5.73 \,(dd, 1H, J = 16.0, 3 \,Hz),$ 5.05-4.63 (m, 1H), 4.37-3.98 (m, 3H), 2.00 (s, 3H), 1.68-1.10 (m, 16H), 0.75 (s, 9H), 0.05 (s, 6H); IR (CCl₄, cm⁻¹) 1715, 1655, 1250; MS (CI⁺) m/e (rel intensity) 401 (M⁺ + 1, 13), 355 (5), 341 (13), 327 (6), 295 (10), 283 (16), 269 (42), 223 (100), 209 (100), 163 (38), 75 (55). 16Za: ¹H NMR (90 MHz, CDCl₃) δ 6.15 (dd, 1H, J = 12.0, 7.5 Hz), 5.70 (dd, 1H, J = 12.0, 1.0 Hz), 5.48–5.20 (m, 1H), 5.12-4.73 (m, 1H), 4.18 (q, 2H, J = 7.0 Hz), 2.05 (s, 3H), 1.70-1.13 (m, 16H), 0.93 (s, 9H), 0.10 (s, 6H); IR (CCl₄, cm⁻¹) 1715, 1645, 1240; MS (Cl⁺) m/e (rel intensity) 401 (M⁺ + 1, 7), 385 (17), 355 (3), 343 (25), 325 (8), 283 (53), 269 (75), 223 (42), 209 (100), 181 (52), 163 (77), 75 (63).

Ethyl (2E/Z,4S,11R)-11-Acetoxy-4-(tert-butyldimethylsiloxy)-2-dodecenoate (16Eb and 16Zb). The procedure described for 16a was followed. Conversion of 15b (1.30 g. 4.33 mmol) gave pure 16Eb (868 mg, 2.1 mmol, 48%) and pure 16Zb (110 mg, 0.27 mmol, 6%), after chromatography (hexane/EtOAc 15:1). 16Eb: ¹H NMR (90 MHz, CDCl₃) δ 6.90 (dd, 1H, J = 16.0, 4.5 Hz), 5.73 (dd, 1H, J = 16.0, 2.0 Hz), 5.15-4.65 (m, 1H), 4.47-4.07 (m, 3H), 2.07 (s, 3H), 1.80-1.20 (m, 18H), 0.98 (s, 9H), 0.13 (s, 6H); IR (CCl₄, cm⁻¹) 1720, 1655, 1245; MS (CI⁺) m/e (rel intensity) 415 (M⁺ + 1, 27), 369 (4), 355 (11), 309 (13), 283 (49), 237 (60), 223 (100), 177 (30), 75 (59). 16Zb: 1H NMR (90 MHz, CDCl₃) δ 6.15 (dd, 1H, J = 12.0, 7.5 Hz), 5.68 (d, 1H, J = 12.0 Hz), 5.50–5.20 (m, 1H), 5.08–4.70 (m, 1H), 4.20 (q, 2H, J = 7.5Hz), 2.07 (s, 3H), 1.80-1.20 (m, 18H), 0.99 (s, 9H), 0.11 (s, 6H); IR (CCl₄, cm⁻¹) 1720, 1645, 1240; MS (CI⁺) m/e (rel intensity) 415 $(M^+ + 1, 3), 399 (3), 357 (20), 297 (100), 283 (18), 237 (11), 223$ (34), 177 (20), 75 (49).

Ethyl (2E/Z,4S,12R)-12-Acetoxy-4-(tert-butyldimethylsiloxy)-2-tridecenoate (16Ec and 16Zc). The procedure described for 16a was followed. Conversion of 15c (2.7 g) gave 3.5 g of crude product. This product was purified by chromatography (petroleum ether/hexane 15:1), giving a fraction of pure 16Ec and a fraction which contained both 16Ec and 16Zc. The latter fraction was again purified by chromatography (petroleum ether (60-80 °C)/EtOAc 20:1), giving the pure isomers. The fractions with 16Ec were merged, giving 1.55 g (3.6 mmol, 42%) of pure product. Compound 16Zc (152 mg, 0.36 mmol) was obtained in 4% yield. 16Ec: ¹H NMR (90 MHz, CDCl₃) δ 6.87 (dd, 1H, J = 15.0, 5.0 Hz), 5.90 (dd, 1H, J = 15.0, 1.5 Hz), 4.83 (sext., 1H), 4.36-4.13 (m, 1H), 4.13 (q, 2H, J = 6.7 Hz), 1.95 (s, 3H), 1.80-1.20(m, 17H), 1.15 (d, 3H, J = 6.0 Hz), 0.86 (s, 9H), 0.02 and 0.00 (s, 3H)6H); IR (CCl₄, cm⁻¹) 1715, 1640, 1245; MS (CI⁺) m/e (rel intensity) $429 (M^+ + 1, 17), 383 (9), 369 (34), 323 (27), 311 (42), 297 (56),$ 251 (74), 237 (100), 191 (21), 75 (48). 16Zc: 1H NMR (90 MHz, CDCl₈) δ 6.17 (dd, 1H, J = 11.0, 7.5 Hz), 5.70 (d, 1H, J = 11.0 Hz), 5.47-5.18 (m, 1H), 4.93 (sext., 1H), 4.23 (q, 2H, J = 7.0 Hz), 2.07 (s, 3H), 1.83-1.13 (m, 20H), 0.93 (s, 9H), 0.12 (s, 3H), 0.07 (s, 3H); IR (CCl₄, cm⁻¹) 1720, 1655, 1240; MS (CI⁺) m/e (rel intensity) 429 (M⁺ + 1, 4), 413 (8), 383 (5), 371 (76), 311 (100), 297 (39), 251 (5), 237 (45), 191 (8), 75 (19).

Saponification of Compound 16Ea with LiOH (General Procedure). LiOH (1.5 mmol, 36 mg) was added to a stirred solution of 16Ea (100 mg, 0.25 mmol) in THF (5 mL) and H₂O (5 mL). This solution was heated at 50 °C for 6 h. (The reaction was monitored by TLC.) The reaction mixture was acidified with tartaric acid to pH 4 and extracted three times with ether. The combined etheral layers were dried (MgSO₄) and concentrated *in vacuo*, giving (2E,4S,10R)-10-hydroxy-4-(*tert*-butyldimethylsiloxy)-2-undecenecarboxylic acid (17Ea) (80 mg, 0.242 mmol) in 97% yield. This product was used without purification in the next step: ¹H NMR (90 MHz, CDCl₃) δ 7.00 (dd, 1H, J = 15.0, 5.2 Hz), 6.55 (bs, 2H), 6.00 (dd, 1H, J = 15.0, 1.5 Hz), 4.47-4.10 (m, 1H), 4.00-3.57 (m, 1H), 1.77-1.10 (m, 10H), 1.20 (d, 3H, J = 6.0 Hz), 0.90 (s, 9H), 0.10 (s, 6H).

(2Z,4S,10R)-10-Hydroxy-4-(*tert*-butyldimethylsiloxy)-2undecenecarboxylic Acid (17Zb). The procedure for 17Ea was followed. From 16Zb (110 mg, 0.266 mmol), 65 mg of crude 17Zb (0.19 mmol, 71%) was obtained after 4 days at 60 °C. This product was immedately used in the text step: ¹H NMR (90 MHz, CDCl₃) δ 6.27 (dd, 1H, J = 11.3, 8.3 Hz), 6.07 (bs, 2H), 5.73 (d, 1H, J = 11.3 Hz), 5.43-5.10 (m, 1H), 4.07-3.63 (m, 1H), 1.77-1.17 (m, 12H), 1.20 (d, 3H, J = 6.0 Hz), 0.93 (s, 9H), 0.13 (s, 3H), 0.10 (s, 3H).

Saponification of 16Eb with NaOH (General Procedure). A solution of NaOH (20 mg, 0.5 mmol) in absolute EtOH (2 mL) was added to a solution of 16Eb (100 mg, 0.241 mmol) in absolute EtOH (5 mL). After 5 h of stirring at 50 °C, the reaction mixture was concentrated, followed by addition of water. This solution was extracted once with ether to remove minor impurities. Then the aqueous layer was acidified with citric acid to pH 4 and extracted three times with ether. The combined organic layers were dried (MgSO₄) and concentrated *in vacuo*, giving (2E,4S,- 11R)-11-hydroxy-4-(*tert*-butyldimethylsiloxy)-2-dodecenecarboxylic acid (17Eb) (82 mg, 0.238 mmol, 99%). This product was without purification used in the next step: ¹H NMR (90 MHz, $CDCl_8$) δ 7.03 (dd, 1H, J = 15.0, 4.50 Hz), 6.03 (dd, 1H, J = 15.0, 2.0 Hz), 5.20 (bs, 2H), 4.53-4.20 (m, 1H), 4.00-3.57 (m, 1H), 1.80-1.20 (m, 12H), 1.27 (d, 3H, J = 6.0 Hz), 0.97 (s, 9H), 0.17 (s, 6H).

(2E,4S,10R)-10-Hydroxy-4-(*tert*-butyldimethylsiloxy)-2undecenecarboxylic Acid (17Za). The synthesis of 17Za was carried out using the procedure described for product 17Eb, giving crude 17Za (0.115 mmol, 99%) after 9.5 h at 78 °C. This product was used in the next step without purification: ¹H NMR (90 MHz, CDCl₃) δ 6.6 (bs, 2H), 6.27 (dd, 1H, J = 12.0, 7.5 Hz), 5.70 (dd, 1H, J = 12.0, 1.0 Hz), 5.40–5.11 (m, 1H), 4.00–3.60 (m, 1H), 1.70–1.11 (m, 10H), 1.20 (d, 3H, J = 7.0 Hz), 0.90 (s, 9H), 0.07 (s, 6H).

(2E,4S,12R)-12-Hydroxy-4-(*tert*-butyldimethylsiloxy)-2tridecenecarboxylic Acid (17Ec). The procedure for the synthesis of 17Eb was followed. From 16Ec (100 mg, 0.23 mmol), 80 mg of crude 17Ec (0.223 mmol, 97%) was obtained after 30 h at 60 °C. The product was immediately used in the next step: ¹H NMR (90 MHz, CDCl₃) δ 7.04 (dd, 1H, J = 15.0, 5.00 Hz), 6.30 (bs, 2H), 6.00 (dd, 1H, J = 15.0, 1.5 Hz), 4.50–4.10 (m, 1H), 4.00– 3.57 (m, 1H), 1.73–1.07 (m, 14H), 1.20 (d, 3H, J = 6.0 Hz), 0.93 (s, 9H), 0.10 (s, 6H).

(2Z,4S,12R)-12-Hydroxy-4-(*tert*-butyldimethylsiloxy)-2tridecenecarboxylic Acid (17Zc). The procedure for the synthesis of 17Eb was followed. From 16Zc (96 mg, 0.224 mmol), 80 mg of 17Zc (0.223 mmol, 99%) was obtained after 24 h at 78 °C. This product was used in the next step without purification: ¹H NMR (90 MHz, CDCl₃) δ 6.37 (bs, 2H), 6.27 (dd, 1H, J = 12.0, 8.0 Hz), 5.73 (d, 1H, J = 12.0 Hz), 5.47-5.10 (m, 1H), 4.05-3.60 (m, 1H), 1.77-1.20 (m, 14H), 1.25 (d, 3H, J = 6.0 Hz), 0.97 (s, 9H), 0.10 (s, 6H).

(2E,4S,10R)-4-(tert-Butyldimethylsiloxy)-2-undecen-10olide (18Ea). To a solution of 17Ea (81 mg, 0.25 mmol) in THF (5 mL) under argon was added Et₃N (0.275 mmol, 28 mg) followed by 2,6-dichlorobenzoyl chloride (0.25 mmol, 52 mg). After the solution was stirred for 2 h, the precipitate was removed by filtration under argon flow. The filtrate was diluted with 130 mL of dry toluene and slowly added (in 2 h) to a refluxing solution of DMAP (181 mg, 1.5 mmol) in toluene. After continued heating for 1 h, the reaction mixture was allowed to cool down to rt and stirring was continued for 16 h. The reaction mixture was diluted with $\approx 100 \text{ mL}$ of ether and washed with saturated tartaric acid and saturated NaHCO₃. The organic layer was dried (MgSO₄) and concentrated in vacuo. The residue was purified by chromatography (hexane/ether 20:1), giving 45 mg of 18Ea (0.144 mmol, 58%): ¹H NMR (400 MHz, CDCl₃) δ 6.50 (dd, 1H, J = 16.1, 8.5 Hz), 6.22 (d, 1H, J = 16.1 Hz), 4.69-4.59 (m, 1H), 4.36-4.28 (m, 1H), 1.76–1.25 (m, 10H), 1.33 (d, 3H, J = 6.2 Hz), 0.88 (s, 9H), 0.05 (s, 3H), 0.01 (s, 3H); IR (CCl₄, cm⁻¹) 1715, 1640, 1250; MS (CI⁺) m/e (rel intensity) 313 (M⁺ + 1, 36), 297 (40), 279 (7), 255 (71), 237 (22), 215 (22), 181 (82), 163 (54), 135 (100), 75 (98).

(2Z,4S,10R)-4-(*tert*-Butyldimethylsiloxy)-2-undecen-10olide (18Za). The procedure for the synthesis of 18Ea was carried out with 17Za (38 mg, 0.115 mmol), giving 11 mg of 18Za (30%), after chromatography (hexane/EtOAc 30:1): ¹H NMR (400 MHz, CDCl₃) δ 5.88 (dd, 1H, J = 11.9, 7.55 Hz), 5.78 (d, 1H, J = 11.9Hz), 5.55-4.96 (m, 1H), 4.81-4.73 (m, 1H), 1.87-1.13 (m, 10H), 1.28 (d, 3H, J = 6.4 Hz), 0.88 (s, 9H), 0.07 (s, 3H), 0.01 (s, 3H); IR (CCl₄, cm⁻¹) 1720, 1640, 1255; MS (Cl⁺) m/e (rel intensity) 313 (M⁺ + 1, 44), 297 (89), 255 (100), 237 (17), 215 (21), 181 (100), 163 (84), 135 (57), 75 (38).

(2E,4S,11R)-4-(*tert*-Butyldimethylsiloxy)-2-dodecen-11olide (18Eb). The procedure for the synthesis of 18Ea was followed. From 17Eb (82 mg, 0.238 mmol) 53 mg of 18Eb (0.163 mmol, 67%) was obtained after chromatography (hexane/ether 20:1): ¹H NMR (90 MHz, CDCl₃) δ 6.63 (dd, 1H, J = 160, 5.6Hz), 6.10 (dd, 1H, J = 16.0, 1.5 Hz), 5.33-4.97 (m, 1H), 4.67-4.40 (m, 1H), 1.93-1.10 (m, 12H), 1.27 (d, 3H, J = 6.0 Hz), 1.00 (s, 9H), 0.13 (s, 6H); IR (CCl₄, cm⁻¹) 1715, 1645, 1250; MS (CI⁺) m/e (rel intensity) 327 (M⁺ + 1, 26), 309 (26), 269 (100), 251 (26), 195 (30), 149 (53), 75 (89).

(2Z,4S,11R)-4-(tert-Butyldimethylsiloxy)-2-dodecen-11olide (18Zb). The procedure for 18Ea was followed. From 17Zb (65 mg, 0.19 mmol), 20 mg of 18Zb (0.061 mmol, 32%) and 17 mg of 18Eb (0.052 mmol, 27%) were obtained after chromatography (hexane/ether 20:1): ¹H NMR (90 MHz, CDCl₃) δ 6.05 (dd, 1H, J = 12.0, 7.80 Hz), 5.73 (d, 1H, J = 12.0 Hz), 5.33-4.93 (m, 2H), 2.17-1.15 (m, 12H), 1.38 (d, 3H, J = 6.0 Hz), 0.99 (s, 9H), 0.17 (s, 3H), 0.14 (s, 3H); IR (CCl₄, cm⁻¹) 1715, 1650, 1255; MS (CI⁺) m/e (rel intensity) 327 (M⁺ + 1, 33), 269 (100), 251 (23), 195 (30), 177 (22), 149 (53), 75 (89).

(2E,4S,12R)-4-(tert-Butyldimethylsiloxy)-2-tridecen-12olide (18Ec). The procedure for the synthesis of 18Ea was followed. From 17Ec (80 mg, 0.223 mmol), 18Ec (21 mg, 0.062 mmol) was obtained in 28% yield after chromatography (hexane/ ether 30:1): ¹H NMR (400 MHz, CDCl₃) δ 6.80 (dd, 1H, J = 15.6, 5.7 Hz), 6.02 (dd, 1H, J = 15.6, 1.3 Hz), 5.18–5.13 (m, 1H), 4.50– 4.42 (m, 1H), 1.80–1.10 (m, 14H), 1.27 (d, 3H, J = 6.4 Hz), 0.89 (s, 9H), 0.06 (s, 3H), 0.04 (s, 3H); IR (CCl₄, cm⁻¹) 1715, 1655; MS (CI⁺) m/e (rel intensity) 341 (M⁺ + 1, 11), 323 (13), 283 (100), 265 (25), 209 (13), 191 (13), 163 (26), 75 (71).

(2Z,4S,12R)-4-(tert-Butyldimethylsiloxy)-2-tridecen-12olide (18Zc). The procedure for the synthesis of 18Ea was followed. From 17Zc (82 mg, 0.23 mmol), 18Zc (16.4 mg, 0.048 mmol) was obtained in 21% yield after chromatography (petroleum ether (60-80 °C)/ether 30:1): ¹H NMR (400 MHz, CDCl₃) δ 5.86 (dd, 1H, J = 12.0, 8.50 Hz), 5.79 (dd, 1H, J = 12.0, 1.0 Hz), 5.28-5.16 (m, 2H), 1.72-1.10 (m, 14H), 1.23 (d, 3H, J = 6.3 Hz), 0.89 (s, 9H), 0.10 (s, 3H), 0.08 (s, 3H); IR (CCl₄, cm⁻¹) 1715, 1640, 1250; MS (CI⁺) m/e (rel intensity) 341 (M⁺ + 1, 51), 325 (16), 283 (100), 265 (23), 209 (47), 191 (18), 163 (22), 75 (45).

(2E,4S,10R)-4-Hydroxy-2-undecen-10-olide (19Ea). n-Bu-NF (0.18 mL, 1 M, in THF) was added via a syringe to a solution of 18Ea (55 mg, 0.18 mmol) in THF at -10 °C under argon. After 3 h of stirring at -10 °C, the reaction mixture was quenched with saturated NH4Cl solution. The mixture was extracted three times with EtOAc. The combined organic layers were dried over $MgSO_4$ and concentrated in vacuo. The residue was purified by chromatography (ether), giving 20 mg of pure 19Ea (0.10 mmol, 56%) as a white solid which was recrystallized from hexane: mp 126–128 °C; $[\alpha]^{20}$ –3.25° (c 0.40, EtOH); ¹H NMR (400 MHz, $CDCl_3$) δ 6.46 (dd, 1H, J = 16.1, 8.9 Hz), 6.34 (d, 1H, J = 16.1 Hz), 4.65-4.54 (m, 1H), 4.45-4.35 (m, 1H), 2.50 (bs, 1H), 2.00-1.35 (m, 10H), 1.34 (d, 3H, J = 6.4 Hz); ¹H NMR (400 MHz, CD_3CN) δ 6.46 (d, 1H, J = 16.2 Hz), 6.23 (dd, 1H, J = 16.2, 9.2 Hz), 4.59-4.53 (m, 1H), 4.30-4.24 (m, 1H), 3.17 (bs, 1H), 1.87-1.13 (m, 10H), 1.26 (d, 3H, J = 6.1 Hz); ¹³C NMR (100 MHz, CDCl₃) & 168.87, 145.70, 122.61, 74.86, 72.39, 35.10, 33.70, 23.48, 23.13, 22.14, 21.24; IR (CCl₄, cm⁻¹) 3590, 3700-3100, 1710, 1655; MS (CI⁺) m/e (rel intensity) 199 (M⁺ + 1, 19), 181 (93), 163 (60), 153 (51), 135 (100), 121 (78), 111 (47), 99 (41), 95 (74), 83 (67), 55 (37); HRMS calcd for C₁₁H₁₆O₂ (M - H₂O) 180.1150, found 180.1151; UV (CH₃CN) λ_{max} 193.9 nm ($\epsilon = 8058 \text{ mol}^{-1} \text{ cm}^{-1}$).

(2Z,4S,10R)-4-Hydroxy-2-undecen-10-olide (19Za). The procedure for the synthesis of 19Ea was followed. From 11 mg of 18Za (0.035 mmol), 5 mg of 19Za (0.025, mmol, 71%) was obtained as a white solid after chromatography (ether): mp 93-95 °C; $[\alpha]^{20}_{\rm D}$ +23.6° (c 0.11, EtOH); ¹H NMR (400 MHz, CDCl₃) δ 5.91 (dd, 1H, J = 12.0, 6.1 Hz), 6.87 (d, 1H, J = 12.0 Hz), 5.05-4.98 (m, 1H), 4.84-4.79 (m, 1H), 2.17 (bs, 1H), 1.78-1.42 (m, 10H), 1.26 (d, 3H, J = 7.7 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 166.82, 143.75, 122.05, 72.15, 67.90, 35.77, 33.84, 26.08, 23.24, 22.44, 20.63; IR (CCl₄, cm⁻¹) 3610, 3600-3150, 1720; MS (CI⁺) m/e (rel intensity) 199 (M⁺ + 1, 6), 181 (22), 163 (26), 135 (23), 101 (100), 83 (48), 55 (35); HRMS calcd for C₁₁H₁₆O₂ (M - H₂O) 180.1150, found 180.1149; UV (CH₃CN) λ_{max} 191.0 nm (ϵ = 1529 mol⁻¹ cm⁻¹), λ 216 nm (ϵ = 716 mol⁻¹ cm⁻¹).

(2E,4S,11R)-4-Hydroxy-2-dodecen-11-olide (19Eb). The procedure for the preparation of 19Ea was used. From 48 mg of 18Eb (0.150 mmol), 22 mg of 19Eb (0.104 mmol, 69%) was obtained as a colorless oil after chromatography (hexane/ether 2:1): $[\alpha]^{20}_{D} + 6.6^{\circ}$ (c 0.42, EtOH); ¹H NMR (90 MHz, CDCl₃) δ 6.82 (dd, 1H, J = 16.5, 6.3 Hz), 6.07 (d, 1H, J = 16.5 Hz), 5.23-4.85 (m, 1H), 4.58-4.33 (m, 1H), 2.03-0.93 (m, 12H), 1.32 (d, 3H, J = 6.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 168.10, 149.69, 121.49, 73.18, 70.89, 35.89, 32.84, 28.26, 27.83, 22.16, 20.74, 19.34; IR (CCL, cm⁻¹) 3610, 3650-3150, 1715, 1645; MS (CI⁺) m/e (rel intensity) 213 (M⁺ + 1, 65), 195 (64), 177 (23), 149 (100); HRMS

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calcd for $C_{12}H_{20}O_3$ 212.1412, found 212.1413; UV (CH₃CN) λ_{max} 207.8 nm (ϵ 7380 mol⁻¹ cm⁻¹).

(2Z,4S,11R)-4-Hydroxy-2-dodecen-11-olide (19Zb). The procedure for the preparation of 19Ea was followed. The deprotection was carried out starting from 20 mg of 18Zb (0.095 mmol). This gave 13 mg of 19Zb (0.061 mmol, 64%) as a white solid after chromatography (hexane/ether 2:1). The product was chromatographed again (petroleum ether (60-80 °C)/ether 2:1) and recrystallized from hexane, giving 5 mg of colorless crystals: mp 75 °C; $[\alpha]^{20}$ +71.7° (c 0.42, EtOH); ¹H NMR (400 MHz, $CDCl_3$) δ 6.02 (dd, 1H, J = 11.9, 8.5 Hz), 5.79 (dd, 1H, J = 11.9, 1.2 Hz), 5.12–5.00 (m, 2H), 1.83 (bs, 1H), 1.68–1.25 (m, 12H), 1.28 $(d, 3H, J = 6.5 Hz); {}^{13}C NMR (100 MHz, CDCl_3) \delta 166.38, 146.80,$ 121.08, 72.18, 66.54, 34.02, 29.64, 24.66, 22.58, 22.37, 20.11, 17.36; IR (CCl₄, cm⁻¹) 3615, 3600-3300, 1710; MS (CI⁺) m/e (rel intensity) 213 (M⁺ + 1, 7), 195 (36), 177 (23), 149 (35); HRMS calcd for C12H20O3 212.1412, found 212.1413; UV (CH3CN) λmax 201.0 nm $(\epsilon = 8967 \text{ mol}^{-1} \text{ cm}^{-1}).$

(2E,4S,12R)-4-Hydroxy-2-tridecen-12-olide (19Ec). The procedure described for 19Ea was followed. From 21 mg of 18Ec (0.062 mmol), 9.3 mg of 19Ec (0.041 mmol, 66%) was obtained as a white solid after chromatography (petroleum ether (60-80 °C)/ether 2:1): mp 49-51 °C; $[\alpha]^{20}_{\rm D}$ +1.33° (c 0.40, EtOH); ¹H NMR (400 MHz, CDCl₃) δ 6.89 (dd, 1H, J = 15.8, 6.9 Hz), 6.02 (dd, 1H, J = 15.8, 1.0 Hz), 5.17-5.05 (m, 1H), 4.53-4.44 (m, 1H), 1.90-1.07 (m, 15H), 1.27 (d, 3H, J = 6.4 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 166.93, 149.38, 121.46, 72.26, 70.81, 34.77, 34.50, 28.44, 28.20, 26.43, 23.25, 20.40, 20.22; IR (CCl₄, cm⁻¹) 3610, 3650-3100, 1715, 1650; MS (CI⁺) m/e (rel intensity) 227 (M⁺ + 1, 66), 209 (55), 191 (19), 163 (100), 149 (23), 111 (35), 84 (46), 69 (44), 55 (52); HRMS calcd for C₁₃H₂₂O₃ 226.1569, found 226.1570; UV (CH₃CN) λ_{max} <190 nm, λ 205.7 nm (ϵ = 10440 mol⁻¹ cm⁻¹), λ 270 nm (ϵ = 595 mol⁻¹ cm⁻¹).

(2Z,4S,12R)-4-Hydroxy-2-tridecen-12-olide (19Zc). Since the conditions used for 19Ea failed to convert 18Zc, the following procedure was used. 18Zc (16 mg, 0.047 mmol) was dissolved in 3:1:1 HOAc/THF/H₂O (1 mL). After overnight stirring, the reaction mixture was extracted three times with ether. The combined organic layers were washed with saturated NaHCO₃, dried (MgSO₄), and concentrated in vacuo. The residue was purified by chromatography (petroleum ether (60-80 °C)/ether 1:1), giving 19Zc (5.3 mg, 0.023 mmol, 49%) as a white solid: mp 44-47 °C; $[\alpha]^{20}$ _D +45.7° (c 0.27, EtOH); ¹H NMR (400 MHz, CDCl₃) δ 5.90 (dd, 1H, J = 11.4, 8.4 Hz), 5.88 (d, 1H, J = 11.4 Hz), 5.26-5.13 (m, 2H), 1.96 (bs, 1H), 1.77-1.12 (m, 14H), 1.24 (d, 3H, J = 6.3 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 166.78, 146.21, 121.93, 71.80, 66.85, 35.18, 34.86, 27.08, 26.67, 25.86, 24.22, 23.63, 20.90; IR (CCl₄, cm⁻¹) 3610, 3650-3100, 1715, 1640; MS (CI⁺) m/e (rel intensity) 227 (M⁺ + 1, 5), 209 (25), 191 (22), 163 (31), 149 (40), 101 (100); HRMS calcd for C13H22O3 226.1569, found 226.1574; UV (CH₃CN) λ_{max} <190 nm, λ 216.0 nm (ϵ = 4318 mol⁻¹ cm⁻¹), λ 280 nm (ϵ = 284 mol⁻¹ cm⁻¹).

Ethyl (2*E*,4*S*)-4-hydroxy-2-nonenoate (20):² ¹H NMR (90 MHz, CDCl₃) δ 6.93 (dd, 1H, J = 16.0, 5.0 Hz), 6.02 (dd, J = 16.0, 1.5 Hz), 4.43-4.13 (m, 3H), 2.00-0.73 (m, 15H); UV (CH₃CN) λ_{max} 207.6 nm (ϵ = 8756 mol⁻¹ cm⁻¹).

Supplementary Material Available: Copies of ¹H NMR spectra for most compounds (60 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.